Contents lists available at ScienceDirect





Science of the Total Environment

#### journal homepage: www.elsevier.com/locate/scitotenv

# Diurnal distribution of airborne bacteria and fungi in the atmosphere of Helwan area, Egypt

A.A. Abdel Hameed <sup>a,\*</sup>, M.I. Khoder <sup>a</sup>, S. Yuosra <sup>a</sup>, A.M. Osman <sup>b</sup>, S. Ghanem <sup>b</sup>

<sup>a</sup> Air Pollution Dept., National Research Centre, Dokki, Giza, Egypt

<sup>b</sup> Botany Dept., Faculty of Science, Helwan Univ., Egypt

#### ARTICLE INFO

Article history: Received 3 July 2009 Received in revised form 17 August 2009 Accepted 24 August 2009 Available online 27 September 2009

*Keywords:* Airborne Fungi Bacteria Diurnal variation Concentration peak

#### ABSTRACT

Airborne bacterial and fungal composition in an industrial town of Helwan, Egypt, was studied using a slit impactor sampler during the period from March 2006 to February 2007. Airborne bacterial concentrations were usually higher than fungi. Bacteria and fungi had similar diurnal variation patterns. Airborne microorganisms reached their concentration peaks in the evening and gradually decreased during the night time. The hourly concentration peaks of the bacteria and fungi appeared at 20:00 h. A significant difference ( $P \le 0.05$ ) was found between the hourly mean concentrations of airborne fungi in winter compared to other seasons. Fungi concentrations were significantly higher ( $P \le 0.05$ ) on working weekdays than weekends. *Aspergillus, Penicillium, Alternaria* and *Cladosporium* were the most predominant airborne fungal genera. *Aspergillus* showed double peak patterns whereas *Penicillium, Alternaria* and *Cladosporium* showed one peak pattern. The diurnal variations of the bacteria and fungi could be divided into four periods: 1) the morning maximum concentration (6:00 h–10:00 h), 2) midday to afternoon pattern (10:00 h–16:00 h), 3) the evening concentration peak (18:00 h–20:00 h) and 4) the gradual decrease of night time concentration (22:00 h–24:00 h). Geographical location, human activity, growth cycle of organisms and meteorological factors were the main criteria controlling the temporal variations of the air microorganisms in the Wadi Hof area.

© 2009 Elsevier B.V. All rights reserved.

#### 1. Introduction

Airborne microorganisms originate from different natural sources such as soil, animals, and humans (Pósfai et al., 2003; Mouli et al., 2005; Fang et al., 2007). Man-made activities such as sewage treatment plants, animal rendering, fermentation processes and agricultural activities emit microorganisms into the air (Recer et al., 2001; Adhikari et al., 2004; Gillum and Levetin, 2008). Airborne microorganisms have been implicated in spoilage of food (Tournas and Katsoudas, 2005), damage of books and archival materials (Aira et al., 2007), biodeterioration of stones (Mohammadi and Krumbein, 2008) and spread of plant and animal diseases (Rossi et al., 2005). Exposure to outdoor air microorganisms has been associated with allergic respiratory symptoms, asthma exacerbation, asthma related death and infection (Dales et al., 2004; Peternel et al., 2004).

Airborne microorganisms have been shown to vary throughout the day and season depending on various environmental factors such as: type of vegetation (Pepeljnjak and Šegvić, 2003), air pollution (Lin and Li, 2000), human activities (Mitakakis et al., 2005), meteorological and seasonal climatic factors (Rossi et al., 2005 and Klarić and Pepeljnjak, 2006) and periodicity of the emission sources (Cariñanos et al., 1999). Members of dry air spora are found in the

greatest abundance in the atmosphere with condition of low humidity and high wind speed (Levetin, 1995). The wet air spora require moisture for release and thus increase in number after precipitation events (Horner et al., 1992).

In Egypt, little attention has been paid to outdoor airborne microorganisms. Airborne fungal concentrations have been monitored in some Egyptian cities including Ismailia (Abdul Wahid et al., 1996), Cairo (Abdel Hameed et al., 1999), Menofia (Abdel Hameed and Khoder, 2001), Western Desert (Ismail et al., 2002), Giza (Abdel Hameed et al., 2007) and New Damietta (EL-Morsy, 2006). Diurnal distribution of airborne fungi was only studied in Upper Egypt, at Assiut (Abu-El-Souod, 1974) and Qena (Abdel-Fattah et al., 1981).

This study aims to assess composition and diurnal distributions of airborne fungi and bacteria on working days and weekends in an industrial district town of Helwan. Moreover the diurnal air distributions of the most common allergenic fungal genera *Alternaria*, *Aspergillus*, *Cladosporium* and *Penicillium* will be determined.

# 2. Materials and methods

#### 2.1. Area description

Air sampling was performed at Wadi Hof site, 29°82′N, 31°33′E, about 4 km north of Helwan district. Helwan is considered one of the major industrial urban areas in Cairo City. It is located at 29°51′N,

<sup>\*</sup> Corresponding author. Tel.: +20 2 33335959; fax: +20 2 33370931. *E-mail address*: hameed\_33@yahoo.com (A.A. Abdel Hameed).

<sup>0048-9697/\$ -</sup> see front matter © 2009 Elsevier B.V. All rights reserved. doi:10.1016/j.scitotenv.2009.08.028



Fig. 1. The mean diurnal concentrations of airborne bacteria and fungi.

31°20′E, about 25 km south of Cairo's city center near the eastern bank of the River Nile with approximately 500,000 inhabitants. Cement, automobiles, iron, steel, textiles, and wood furniture are the main industry products in Helwan area. A variety of vegetation is present in the area but there is no predominant ground cover.

# 2.2. Air sampling and isolation methods

Airborne bacteria and fungi samples were collected during the period from March 2006 to February 2007, at Wadi Hof, Helwan district. Sampling was conducted from the roof of a ~15 meter height building. Slit sampler (*Model*  $\Pi$  818N°5587, CAEIIAHO, BCCCP) was used to collect airborne microorganisms. Airborne particles impact onto a rotating agar plate by passing through a slit at a flow rate of 20 l/min for 2–4 min. Air samples were collected on Mondays and Fridays (weekends), 2 times per month, from 6:00 h to 24:00 h, at two hour intervals. Two consecutive samples were collected during every



Fig. 2. Seasonal diurnal variations of airborne bacteria and fungi.

sampling event (a total of 4 sample runs were collected every month). Petri plates, (in replicates) containing nutrient agar and Rosebengal streptomycin agar media were used to collect airborne bacteria and fungi, respectively. The slit sampler was sterilized by 70% ethanol between each sampling run. Collected samples were maintained in an ice box during sampling and transportation.

Bacterial plates were incubated at 25 °C for 48 h; whereas fungal plates were incubated at 28 °C for 5–7 days and examined daily. The growing colonies were counted and the mean count from replicate plates was calculated. The airborne bacterial and fungal concentrations are calculated and expressed as colony forming unit per cubic meter of air (CFU/m<sup>3</sup>).

Identification of all fungal isolates was done using macroscopic and microscopic features following to Raper and Fennell (1965), Ellis (1971), Singh et al. (1991) and Barnett and Hunter (1999).

#### 2.3. Meteorological parameters

Temperature and relative humidity were measured with a Psychrometer (SATO, PC-5000 TRH-II sampler) during each sampling event. Wind speed records were obtained from the Egyptian Meteorological Authority. During this study, temperature ranged between 12 and 43 °C, relative humidity ranged between 23 and 69.5% and wind speed records varied between 0.5 and 7.2 m/s.

#### 2.4. Statistical analysis

The Mann–Whitney *U* test was used to ascertain the significance of differences between the hourly mean concentrations of airborne microorganisms and on working weekdays and weekends. A probability of less or equal to  $P \le 0.05$  was considered significant.

### 3. Results

# 3.1. The diurnal distributions of airborne bacteria and fungi

Fig. 1 shows the diurnal distributions of airborne bacteria and fungi. The hourly concentrations of bacteria exceeded fungi. The concentration peaks of airborne bacteria  $(1.414 \times 10^3 \text{ CFU/m}^3)$  and fungi  $(5.90 \times 10^2 \text{ CFU/m}^3)$  were achieved at 20:00 h. Seasonal diurnal distributions of the bacteria and fungi are shown in Fig. 2(a–d). Their mean concentration peaks were recorded at 20:00 h in spring, summer and autumn. In contrast, the mean concentration peaks of airborne bacteria  $(1.801 \times 10^3 \text{ CFU/m}^3)$  and fungi  $(2.81 \times 10^2 \text{ CFU/m}^3)$  were observed at 18:00 h in winter, 2 h earlier than other seasons. Statistically, Mann–Whitney *U* test showed significant differences between diurnal distributions of bacteria in spring and summer seasons (*P*=0.0315). Significant differences were found between diurnal distributions of fungi in spring and winter (*P*=0.0262) and in autumn and winter (*P*=0.0002).

# 3.2. Diurnal distributions of airborne bacteria and fungi on working days and weekends

Airborne bacteria and fungi showed similar distribution patterns on working days and weekends (Fig. 3a and b). A non-significant difference was found between mean concentrations of airborne bacteria on working days and weekends (P=0.5147). The concentration peaks of bacteria were achieved at 18:00 h and 20:00 h on working days and weekends, respectively. The concentration peaks of the fungi on working days and weekends were detected at 20:00 h. A significant difference was found between the concentrations of airborne fungi (P=0.0446) on working and weekends, where higher airborne microbial concentrations shifted toward working days.



Fig. 3. Diurnal variations of airborne bacteria (a) and fungi (b) on working days and weekend days.

#### Table 1

The types, number of isolates and percentages of airborne fungal genera identified in Wadi Hof.

| Fungal genera          | Number of isolates | %      |
|------------------------|--------------------|--------|
| Total Aspergillus      | 3116               | 30.84  |
| Aspergillus flavus     | 525                | 5.2    |
| Aspergillus niger      | 1296               | 12.82  |
| Aspergillus fumigatus  | 4                  | 00.04  |
| Aspergillus ochraceous | 43                 | 00.43  |
| Other Aspergillus spp. | 1248               | 12.35  |
| Acremonium             | 2                  | 0.02   |
| Alternaria             | 913                | 09.03  |
| Aureobasidium          | 124                | 01.227 |
| Basidiomycetes         | 9                  | 00.09  |
| Bipolaris              | 12                 | 00.118 |
| Botrytis               | 1                  | 0.001  |
| Botryotricum           | 1                  | 0.001  |
| Chlamydomycetes        | 10                 | 00.1   |
| Cladosporium           | 2575               | 25.51  |
| Curvularia             | 2                  | 00.02  |
| Drechslera             | 5                  | 00.05  |
| Emericella             | 21                 | 00.2   |
| Epicoccum              | 4                  | 00.04  |
| Eurotium               | 6                  | 00.06  |
| Fusarium               | 84                 | 0.831  |
| Geotrichum             | 3                  | 0.029  |
| Monilia                | 5                  | 00.05  |
| Mucor                  | 12                 | 0.118  |
| Mycelia sterile        | 811                | 08.03  |
| Nigrospora             | 3                  | 00.03  |
| Oidiodendron           | 386                | 03.82  |
| Penicillium            | 1733               | 17.15  |
| Rhizopus               | 98                 | 0.969  |
| Stachybotrys           | 9                  | 00.09  |
| Scopulariopsis         | 14                 | 0.138  |
| Torula                 | 3                  | 0.029  |
| Ulocladium             | 14                 | 0.138  |
| Yeast                  | 128                | 01.27  |
| Total isolates         | 10,104             | 100    |

# 3.3. Diurnal distribution of the most predominant airborne fungi

The identified airborne fungi are shown in Table 1. A total of 10,104 fungal isolates belonging to 29 fungal genera were identified. Aspergillus (30.84%), Alternaria (9.03%), Cladosporium (25.51%) and Penicillium (17.15%) were the most common airborne fungi. The diurnal distributions of Alternaria, Aspergillus, Cladosporium, and Penicillium are shown in Fig. 4(a–b). The concentration peak of Alternaria  $(8.7 \times 10 \text{ CFU/m}^3)$ was achieved at 16:00 h whereas the minimum concentration (6.38 CFU/ m<sup>3</sup>) was detected at 10:00 h. Aspergillus showed double peak patterns at 10:00 h  $(1.15 \times 10^2 \text{ CFU/m}^3)$  and at 20:00 h  $(1.44 \times 10^2 \text{ CFU/m}^3)$ . The concentration peaks of Penicillium (9×10 CFU/m<sup>3</sup>) and Cladosporium  $(1.96 \times 10^2 \text{ CFU/m}^3)$  were found at 20:00 h. On the other hand, the minimum concentrations of *Penicillium*  $(2.5 \times 10 \text{ CFU/m}^3)$  and *Cladosporium*  $(3.1 \times 10 \text{ CFU/m}^3)$  were found at 14:00 h. Aspergillus drought appeared 2 h earlier than Penicillium and Cladosporium. Significant differences were found between diurnal concentrations of Aspergillus with Penicillium (P<0.0001) and Alternaria (P<0.0001) and between Alternaria with *Penicillium* (P = 0.0376) and *Cladosporium* (P = 0.0216).

# 4. Discussion

The hourly distribution of biological particles in the atmosphere of a city is analyzed to know at what time of the day the concentration is maximum. This knowledge can be useful for all people who suffer from respiratory diseases and help plan their outdoor activities. In the present study the diurnal variations of airborne bacteria and fungi could be divided into four periods: 1) the morning concentration peak, 6:00 h-10:00 h, 2) midday to afternoon pattern, 10:00 h-16:00 h, 3) the evening concentration peak achieved between 18:00 h and 20:00 h, and 4) the nighttime decreased concentration, 22:00 h-00:00 h. The morning concentration peak may be attributed to the heat of the sun. At sunrise, wind speed and temperature concurrently increase and relative humidity decreases which leads to evaporation of water molecules binding the microbial particles with different surfaces. High wind speed releases microorganisms into the air. In early morning, human activities and traffic flow disturb and resuspend soil particles into air that may increase airborne microbial concentrations. At midday and afternoon higher temperature, intensive solar radiation and lower relative humidity collectively decrease airborne microbial concentrations. Changed water content occurs for all bioaerosol particles and represents the most fundamental potential stress on air microorganisms. Moreover, radiation effect is exacerbated by dehydration (Cox and Wathes, 1995). The decrease in temperature, solar radiation and increase of human activities are the main causes for the evening peak. In addition, increased humidity in the evening helps physical repair of damaged air microorganisms. However, the absence of human activities leads to decrease in



Fig. 4. Diurnal variations of the most common airborne fungi.

airborne microbial concentrations at night time. Stable conditions at night cause fast settlement of air microorganisms which re-suspend again into the air with the sun heat.

The results in the present study agree with Lighthart and Shaffer (1995) who found sunrise peak concentration of bacteria between 06:00 and 08:00 h, which is concurrent with increased winds near the ground level. Fang et al. (2007) observed higher airborne bacterial concentrations at 09:00 and 17:00 h than at 13:00 h due to heavy human activities and traffic flow in the morning. The stability of airborne bacterial patterns between 10:00 h and 16:00 h could be explained by the fact that as the day proceeds, airborne bacteria generated by the flux from vegetation, soil and anthropogenic sources are trapped below the inversion layer and accumulated with time during the day. Concentration gradients were absent and no upward flux occurred due to atmospheric stagnation and presence of thermal inversion in midday period (Lighthart and Shaffer, 1995). Moreover, Lindemann and Upper (1985) found the evening peak at 20:00 h.

Diurnal variations of airborne fungi depend on locality, human activity and weather conditions. Diurnal distributions of airborne fungal spores are associated with the sun heat effect in the morning and reduced relative humidity (Royes, 1987). Nussbaum (1990) reported that morning was the time of day where maturation and dispersion occurred for many fungi. He also attributed fewer airborne fungal spores during midday and afternoon to the effect of higher temperature, and solar radiation. Jones and Harrison (2004) attributed the rising of airborne fungal concentration in the morning to the increasing of wind speed. Fang et al. (2005) explained the morning maximum peak of fungi to the effect of sunlight which causes spore release. In contrast, extreme midday conditions reduce concentrations of biological materials (Jones and Harrison, 2004). Levetin (1995) concluded that hazard of desiccation was the greatest during midday. Concentrations of airborne fungi were lower during daytime than at nighttime due to the effect of solar radiation whereas dark and high relative humidity enhances fungal viability (Fengxiang et al., 1991). Burch and Levetin (2002) recorded two airborne fungal spore peaks, at 08:00 h and at 18:00 h, the authors explained the evening peak as a result of increasing wind speed.

The winter's concentration peak appeared 2 h earlier than other seasons. This is attributed to driving mainly within the city and returning back home soon before the sunset. During other seasons, concentration peaks were delayed until 20:00 h. This may be attributed to the lifestyle of people, higher number of daylight hours and longer staying outdoors.

In this study airborne microorganisms had similar diurnal rhythms on working days and weekends. No significant difference was found between diurnal distributions of airborne bacteria on working days and weekends, probably because of the same lifestyle of people. In contrast, there was significant difference between airborne fungi (P=0.0446) on working days and weekends, because fungal diurnal variations are mainly related to weather conditions throughout the day. The slight differences observed throughout the day may be related to changes in meteorological factors including wind direction and accidental human activity near the sampling location. Fang et al. (2007) reported that atmospheric microbial pollutants are potentially correlated to population density and activity.

Airborne fungi are among the most common organisms in nature and they are considered to be correlated with adverse health effects of humans and plants (Shelton et al., 2002). In this study *Aspergillus*, *Alternaria*, *Cladosporium* and *Penicillium* were the predominant fungal genera in the atmosphere and their concentrations appeared to differ from place to place and even from hour to hour because of prevailing local environment, fungal genera and human activity. *Aspergillus* had double peak patterns, the concentration peaks of *Penicillium* and *Cladosporium* were achieved in the evening at 20:00 h whereas *Alternaria* had a midday peak pattern. These findings agree with Sreeramulu and Ramalingam (1966) who found two daytime maxima of *Aspergillus*, *Helminthosporium* and *Tetrapolea*. Lin and Li (2000) attributed insignificant diurnal pattern for *Aspergillus* and *Penicillium* to their passive launching mechanism of dry small conidia that can be liberated even by slight air currents or vibration. Ho et al. (2005) found the highest concentration of *Aspergillus/Penicillium* around midnight. Fang et al. (2005) found the lowest concentration of *Penicillium* at 13:00 h. Gillum and Levetin (2008) observed the highest concentration of *Penicillium/Aspergillus* spores at 10:00 h.

In this study, Cladosporium peak shifted toward the evening (20:00 h) and gradually increased in the afternoon period between 16:00 h and 20:00 h with the lowest concentration at 14:00 h. A significant concentration peak of Alternaria was recorded at 16:00 h. Cladosporium and Alternaria are usually found in higher concentrations during warmer periods of the day due to their dry-weather spores. The late afternoon peak coincided with the results obtained by Mediavilla (1995) for Cladosporium spores. Padys et al. (1969) found a single *Cladosporium* peak throughout the day occurring at different hours depending on the climatic features of the area. Cladosporium showed midday pattern (10:00 h-16:00 h) and double peak pattern (08:00 h-10:00 h, 14:00 h-18:00 h), (Lacey, 1981). Nayak et al. (1998) observed the *Cladosporium* peak during morning hours rather than at noon and in the evening. Fang et al. (2007) detected the highest concentrations of Cladosporium at 17:00 h. Troutt and Levetin (2001) found the peak concentration of *Cladosporium* at 14:00 h and the lowest at 6:00 h.

*Alternaria* is a member of dry air spora which favors high temperature and low relative humidity at midday. The presence of melanin pigment protects conidia from the impact effect of sunlight and UV radiation (Gregory, 1973). Members of dry-air spores are found in the greatest abundance in the atmosphere characterized by low humidity during warmer afternoon hours (Levetin, 1995). Dry spore types are released into the air during the day whereas wet spores are released at night (Sarica et al., 2005). The present results agree with Corden and Millington (2001) who found a maximum peak of *Alternaria* between 14:00 h and 22:00 h and more than half the daily *Alternaria* catch was caught between 18:00 h and 24:00 h. Sarica et al. (2007) found higher concentrations of *Alternaria* in the evening time. Generally concentration peaks of airborne fungi have varied among various studies all over the world due to differences in human activities, climatic conditions, vegetation and topographical factors.

#### 5. Conclusion

Airborne bacterial and fungal concentrations varied from hour to hour throughout the day. The rhythmic distributions in microbial liberation depend on geographical characters, human activities, meteorological factors and growth cycle of organisms. The concentration peak in winter season appeared 2 h earlier than other seasons. Airborne microorganisms had the same diurnal rhythms on working days and weekends. *Aspergillus, Alternaria, Cladosporium* and *Penicillium* were the dominant fungal genera in the atmosphere. *Aspergillus* showed double peak patterns at 10:00 h and 20:00 h. The concentration peaks of total airborne fungi, bacteria, *Penicillium* and *Cladosporium* appeared at 20:00 h. This time is considered dangerous for people who are suffering from respiratory diseases whereas midday is dangerous for persons who are sensitive to *Alternaria*.

#### References

Abdel Hameed A, Khoder M. Suspended particulates and bioaerosols emitted from an agricultural non-point source. J Environ Monit 2001;3:206–9.

Abdel Hameed A, Farag S, El Abagy M, Mansour F. Airborne Gram negative bacteria in air at three occupational sites in greater Cairo, Egypt. J Microbiol 1999;34:583–94.Abdel Hameed A, Khoder M, Emad A. Fertile fungal spores collected on different faced

surfaces in the atmosphere of Giza, Egypt. Aerobiologia 2007;23:47–57. Abdel-Fattah H, Moubasher A, Swelim M. Studies on air-borne fungi at Qena. II. Diurnal

Abdel-Fattah H, Moubasher A, Swelim M. Studies on air-borne fungi at Qena. II. Diurna fluctuations. Z Allg Mikrobiol 1981;21:177–9. Abdul Wahid A, Moustafa A, Moustafa A. Fungal population in the atmosphere of Ismailia City. Aerobiologia 1996;12:249–55.

Abu-El-Souod S. Studies on fungus air-spora in Egypt. Ph.D. Thesis, Fac. Sci., Assiut Univ 1974.

- Adhikari A, Reponen T, Lee S, Grinshpun S. Assessment of human exposure to airborne fungi in agricultural confinements: personal inhalable sampling versus stationary sampling. Ann Agric Environ Med 2004;11:269–77.
- Aira M, Jato V, Stchigel A, Rodríguez-Rajo F, Piontelli E. Aeromycological study in the Cathedral of Santiago de Compostela (Spain). Int Biodeterior Biodegrad 2007;60: 231–7.
- Barnett H, Hunter B. Illustrated genera of imperfect fungi. 4th ed. St. Paul, MN: The American Phytopathological Society, APS; 1999. p. 218.
- Burch M, Levetin E. Effects of meteorological conditions on spore plumes. Int J Biometeorol 2002;46:107–17.
- Cariñanos P, Galán C, Alcazar P, Dominguez E. Diurnal variation of biological and nonbiological particles in the atmosphere of Córdoba, Spain. Aerobiologia 1999;15: 177–82.
- Corden J, Millington W. The long-term trends and seasonal variation of the aeroallergen *Alternaria* in Derby, UK. Aerobiologia 2001;17:127–36.
- Cox C, Wathes C. Bioaerosols handbook. 1995, CRC Press LLC, Lewis Publishers.
- Dales R, Cakmak S, Judek S, Dann T, Coates F, Brook J, Burnett R. Influence of outdoor aeroallergens on hospitalization for asthma in Canada. J Allergy Clin Immunol 2004;113:303–6.
- Ellis M. Dematiaceous hyphomycetes, 1971; (p.608). The Western Press Ltd: London and Reading Commonwealth Mycological Institute Kew, Surrey, UK.
- EL-Morsy E. Preliminary survey of indoor and outdoor airborne microfungi at coastal buildings in Egypt. Aerobiologia 2006;22:197–210.
- Fang Z, Ouyang Z, Hu L, Wang X, Zheng H, Lin X. Culturable airborne fungi in outdoor environments in Beijing, China. Sci. Total Environ. 2005;350:47–58.
- Fang Z, Ouyang Z, Zheng H, Wang X, Hu L. Culturable airborne bacteria in outdoor environments in Beijing, China. Microbial Ecol 2007;54:487–96.
- Fengxiang C, Qingxuang H, Zhensheng C, Lingyin M, Shigang Y. Factors of influence on microbial pollution in the atmosphere over Beijing area. Aerobiologia 1991;7: 136–43.
- Gillum S, Levetin E. The air spora close to a compost facility in Northeast Oklahoma: part I: spore trap sampling. Aerobiologia 2008;24:3-12.
- Gregory P. The microbiology of the atmosphere. 2nd edn. London: Leonard Hill; 1973. Ho H, Rao C, Hsu H, Chiu Y, Liu C, Chao H. Characteristics and determinants of ambient
- fungal spores in Hualien, Taiwan. Atmos Environ 2005;39:5839–50. Horner W, O'Neil C, Lehrer S. Basidiospore aeroallergens. Clin Rev Allergy 1992;10: 191–211
- Ismail M, Abdel-Hafez S, Moharram A. Aeromycobiota of western desert of Egypt. Afr J Sci Technol (AJST) Sci Eng Ser 2002;3:1–9.
- Jones A, Harrison M. The effects of meteorological factors on atmospheric bioaerosol concentrations—a review. Sci Total Environ 2004;326:151–80.
- Klarić M, Pepeljnjak S. Year-round aeromycological study in Zagreb area, Croatia. Ann Agric Environ Med 2006;13:55–64.
- Lacey J. The aerobiology of conidial fungi. In: Kendrick B, editor. Biology of conidial fungi, Vol. 1. Academic Press; 1981. p. 373–416.
- Levetin E. Fungi. In: Burge HA, editor. Bioaerosols. Boca Raton, Fla: CRC; 1995. p. 87-120. Lighthart B, Shaffer B. Airborne bacteria in the atmospheric surface layer: temporal distribution above a grass seed field. Appl Environ Microbiol 1995;61:1492–6.
- Lin W, Li C. Associations of fungal aerosols, air pollutants, and meteorological factors. Aerosol Sci Tech 2000;32:359–68.

- Lindemann J, Upper C. Aerial dispersal of epiphytic bacteria over bean plants. Appl Environ Microbiol 1985;50:1229–32.
- Mediavilla A. Modelos de variación intradiurna y estacional de la concentración en la atmósfera del Género *Cladosporium*. 1995; Thesis Doctoral. University of Córdoba.
- Mitakakis T, O'Meara T, Tovey E. The effect of sunlight on allergen release from spores of the fungus *Alternaria*. Grana 2005;42:43–6. Mohammadi P, Krumbein W. Biodeterioration of ancient stone materials from the
- Persepolis monuments (Iran). Aerobiologia 2008;24:27–33.
- Mouli C, Mohan S, Reddy S. Assessment of microbial (bacteria) concentrations of ambient air at semi-arid urban region: influence of meteorological factors. AEER 2005;3:139–49.
- Nayak B, Nanda A, Behera N. Airborne fungal spores in an industrial area: seasonal and diurnal periodicity. Aerobiologia 1998;14:59–67.
- Nussbaum F. Variation in the airborne fungal spore population of the Tuscarawas Valley with respect to microenvironment, time of day, and date. OHIO J Sci 1990;90:77–86.
- Padys S, Kramer C, Clary R. Periodicity in spore release in *Cladosporium*. Mycologia 1969;61:87–98.
- Pepeljnjak S, Šegvić M. Occurrence of fungi in air and on plants in vegetation of different climatic regions in Croatia. Aerobiologia 2003;19:11–9.
- Peternel R, Čulig J, Hrga I. Atmospheric concentrations of *Cladosporium* spp. and *Al-ternaria* spp. spores in Zagreb (Croatia) and effects of some meteorological factors. Ann Agric Environ Med 2004;11:303–7.
- Pósfai M, Li J, Anderson J, Buseck P. Aerosol bacteria over the Southern Ocean during ACE-1. Atmos Res 2003;66:231–40.
- Raper K, Fennell D. The genus Aspergillus. Baltimore, USA: The Williams and Wilkins Comp; 1965. p. 686.
- Recer G, Browne M, Horn E, Hill K, Boehler W. Ambient air levels of *Aspergillus fumigatus* and thermophilic actinomycetes in a residential neighborhood near a yard-waste composting facility. Aerobiologia 2001;17:99-108.
- Rossi V, Bugiani R, Giosué S, Natali P. Patterns of airborne conidia of *Stemphylium vesicarium*, the causal agent of brown spot disease of pears, in relation to weather conditions. Aerobiologia 2005;21:203–16.
- Royes J. Some components of the airspora in Jamaica and their possible medical application. Grana 1987;26:151–7.
- Sarica S, Asan A, Tungan Y, Ture M. Airborne fungal concentrations in east patch of Edirne City (Turkey) in autumn using two sampling methods. Trakya Univ J Sci 2005;6:97-106.
- Sarica S, Asan A, Sabuncuoğlu Y, Yavuz E. Airborne fungal concentrations of morning and evening in east patch of Edirne City using two sampling methods. Trakya Univ J Sci 2007;8:15–20.
- Shelton B, Kirkland K, Flanders W, Morris G. Profiles of airborne fungi in buildings and outdoor environments in the United States. Appl Environ Microbiol 2002;68:1743–53.
- Singh K, Frisvad J, Thrane U, Mathur S. An illustrated manual on identification of some seed-borne Aspergilli, Fusaria, Penicillia and their mycotoxins. Danish Government Institute of Seed Pathology for Developing Countries, Ryvangs Alle 78 DK-2990 Hellerup: Denmark; 1991.
- Sreeramulu T, Ramalingam A. Two-year study of the air spora of a paddy field near Visakhapatnam. Ind J Agric Sci 1966;36:111–32.
- Tournas V, Katsoudas E. Mould and yeast flora in fresh berries, grapes and citrus fruits. Int J Food Microbiol 2005;105:11–7.
- Troutt C, Levetin E. Correlation of spring spore concentrations and meteorological conditions in Tulsa, Oklahoma. Int. J. Biometeorol 2001;45:64–74.